

In vivo multiphoton imaging of a filamentous fungus *Phycomyces blakesleeanus*: the effect of small ambient temperature increase on mitochondrial morphology and lipid droplets density

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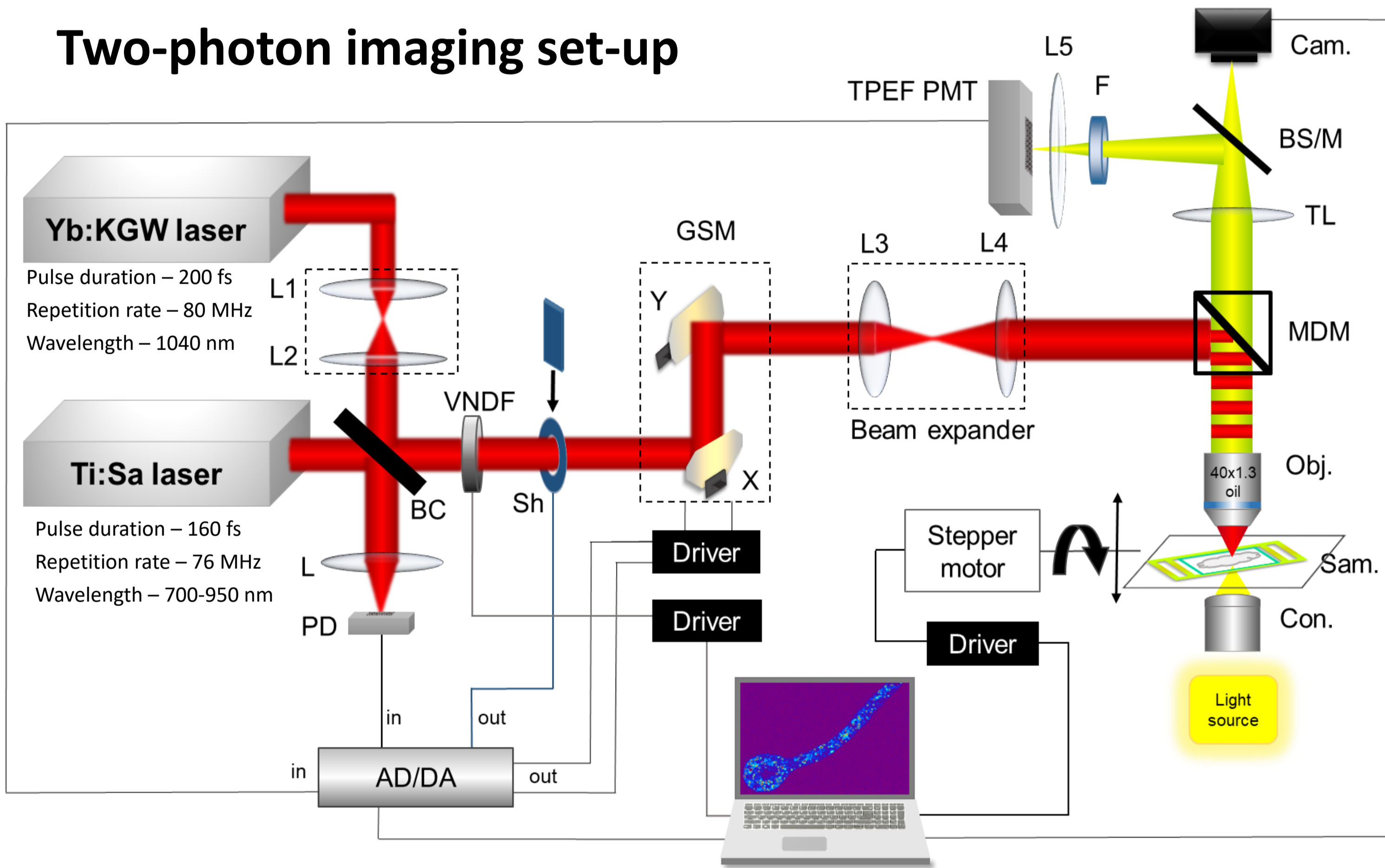


Introduction

Mitochondrial function, and consequently cellular metabolic status and fitness of a cell, is tightly linked to the dynamic changes of mitochondrial morphology, including mitochondrial fusion, fission and mitophagy [1]. Lipid droplets (LDs) can be in close contact with mitochondria, and accumulate autophagy or mitophagy generated material during the reparatory processes [2]. The effect of increased ambient temperature on mitochondrial morphology and LDs density in living cells of the filamentous fungus *Phycomyces blakesleeanus* was investigated.

For *in vivo* imaging of mitochondria and lipid droplets **two-photon excitation fluorescence (TPEF) microscopy** was used.

Two-photon imaging set-up

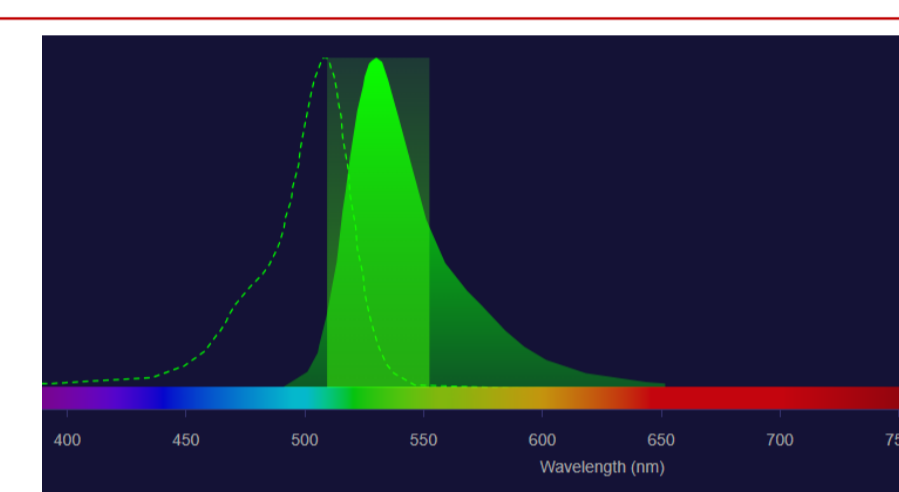


Schematic drawing of NLSM setup. L1 and L2 - lenses of 1:1 beam expander for recollimation, BC - beam combiner, VNDF - motorized variable neutral density filter, Sh - shutter, GSM - galvanometer-scanning mirrors, L3 and L4 - lenses of 1:3.75 beam expander, MDM - main dichroic mirror (cut-off 700 nm), Con. - aspheric condenser lens, TL - tube lens, BS/M - beam splitter or mirror toggle, F - VIS filter 400-700 nm + bandpass interference filter 530/43 (for Rh123) or 570LP (NR), L5 - focusing lens, TPEF PMT - photomultiplier tube for TPEF signal, L - lens, PD - photodiode, AD/DA - acquisition card.

Advantages:

- High contrast images
- 3D imaging in high resolution
- Reduced photodamage and photobleaching of the sample using IR ultrafast pulsed lasers
→ possibility of extended *in vivo* imaging

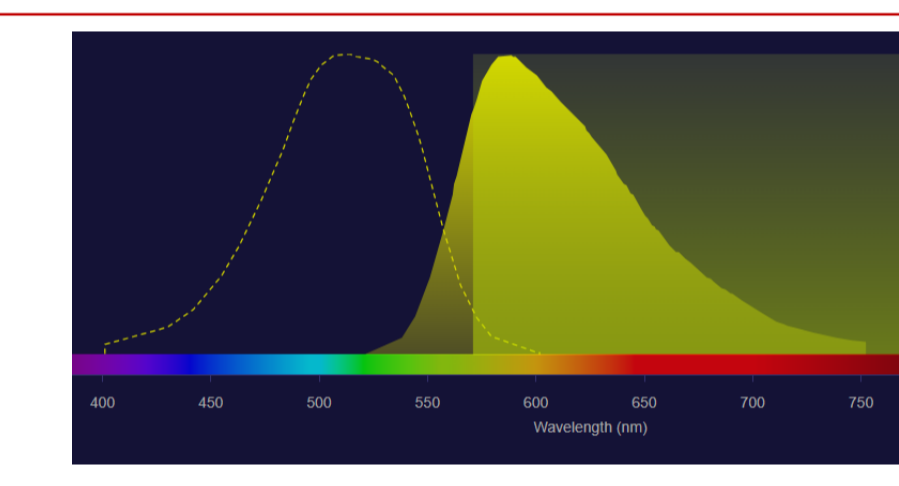
Two-photon exc. (800 nm) of Rhodamine 123



Rhodamine 123 stains active mitochondria in living cells. Dye entry depends on the mitochondrial membrane potential.

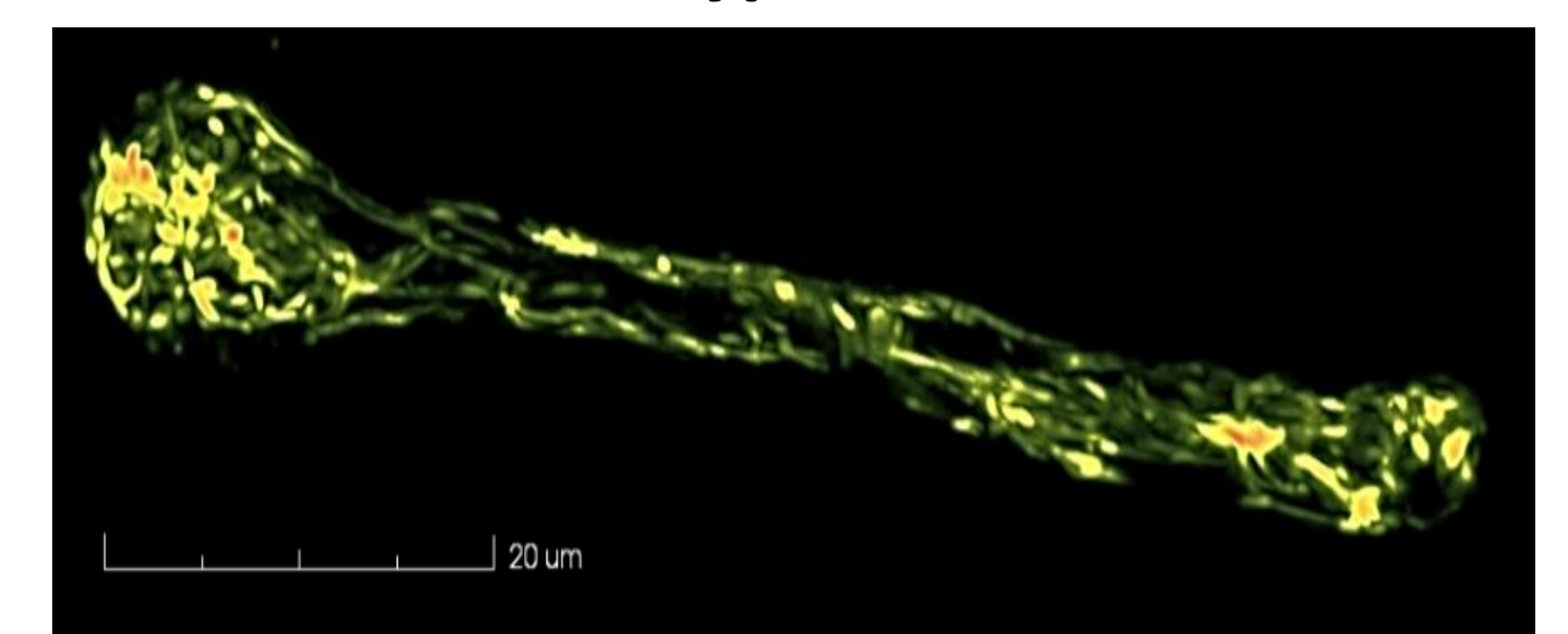
Vital dyes

Two-photon exc. (1040 nm) of Nile Red dye

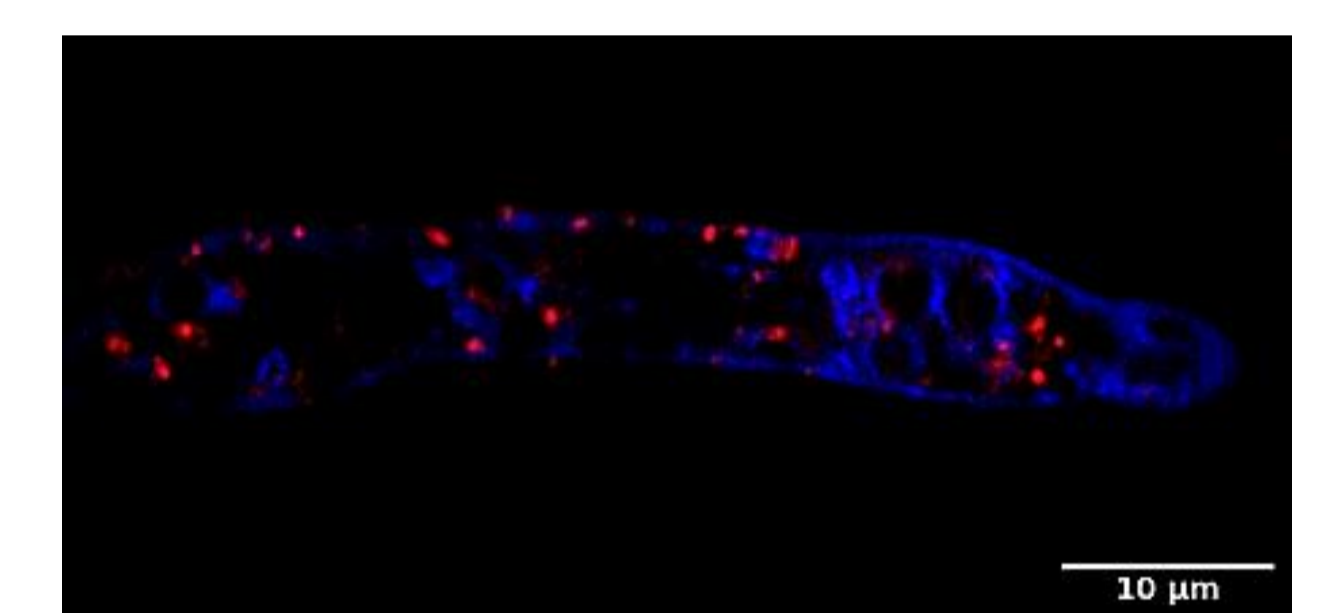


Nile Red stains cellular lipid droplets

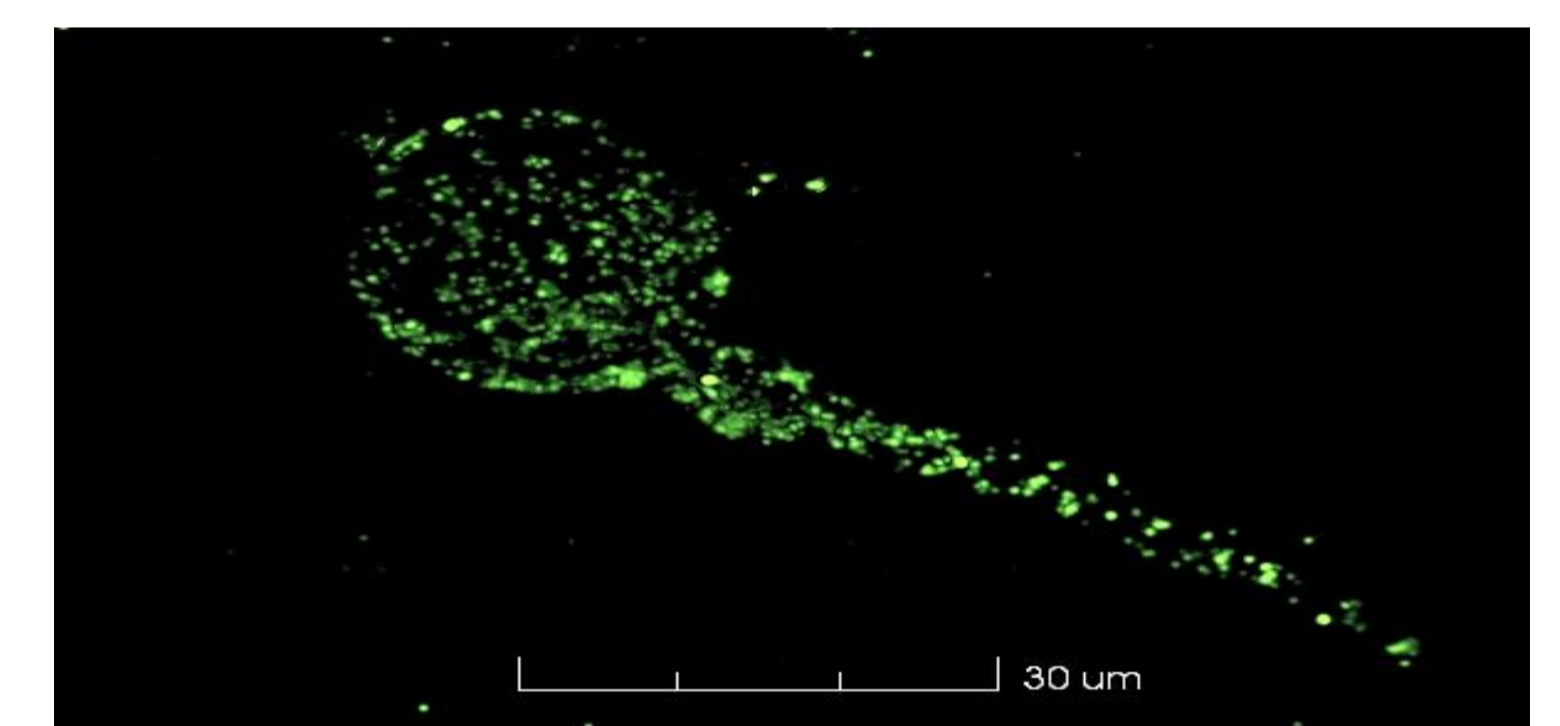
TPEF imaging of *Phycomyces blakesleeanus* live hyphae



3D model (21 slices/images, 0.9 μm apart along the z axis) of tubular mitochondria (22°C) in live hypha

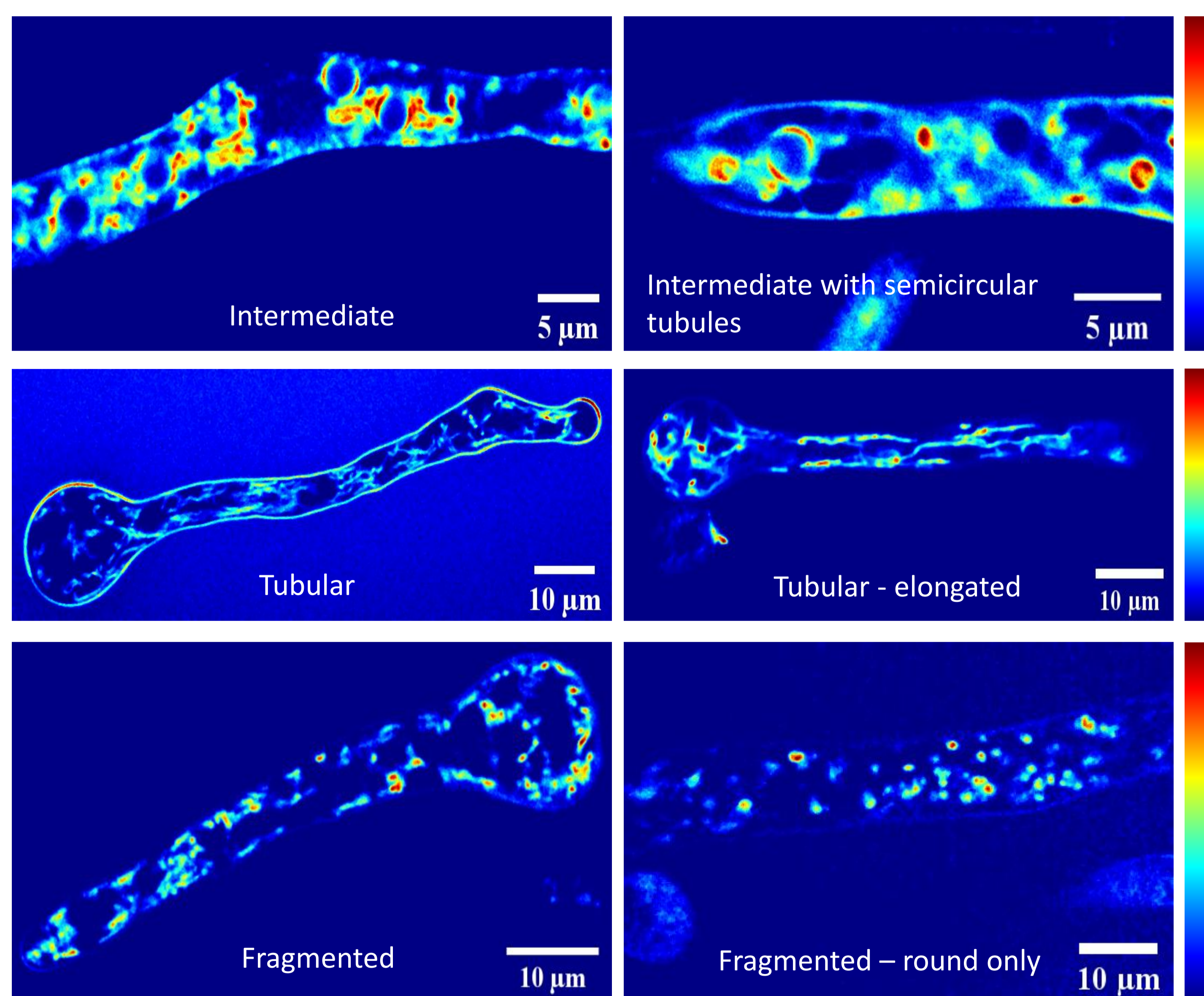


Merged 2D image of Rh123 (blue signal) and NR (red signal) in same hypha



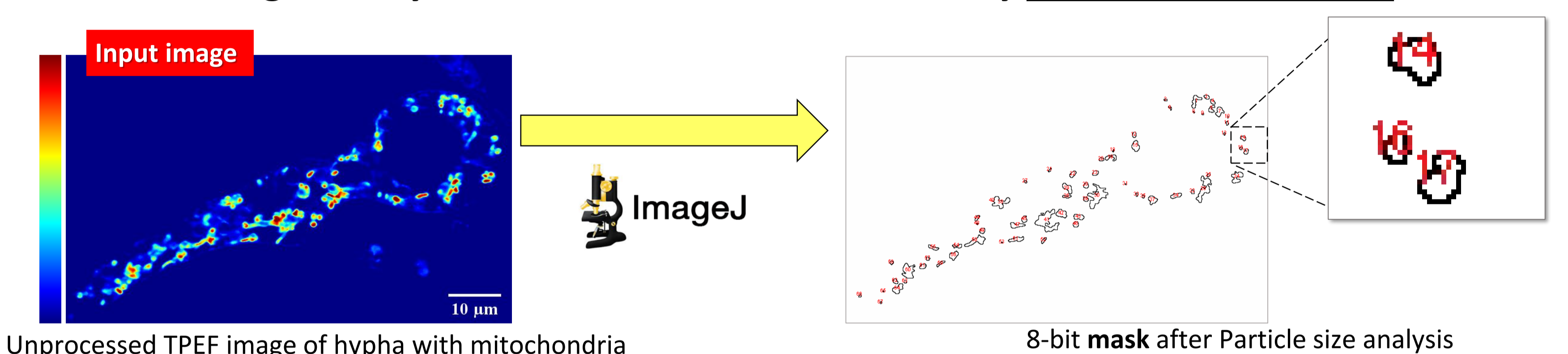
3D model (15 slices/images, 0.9 μm apart along the z axis) of lipid droplets (25°C) in live hypha

TPEF images of mitochondrial morphology in *Phycomyces blakesleeanus* hyphae

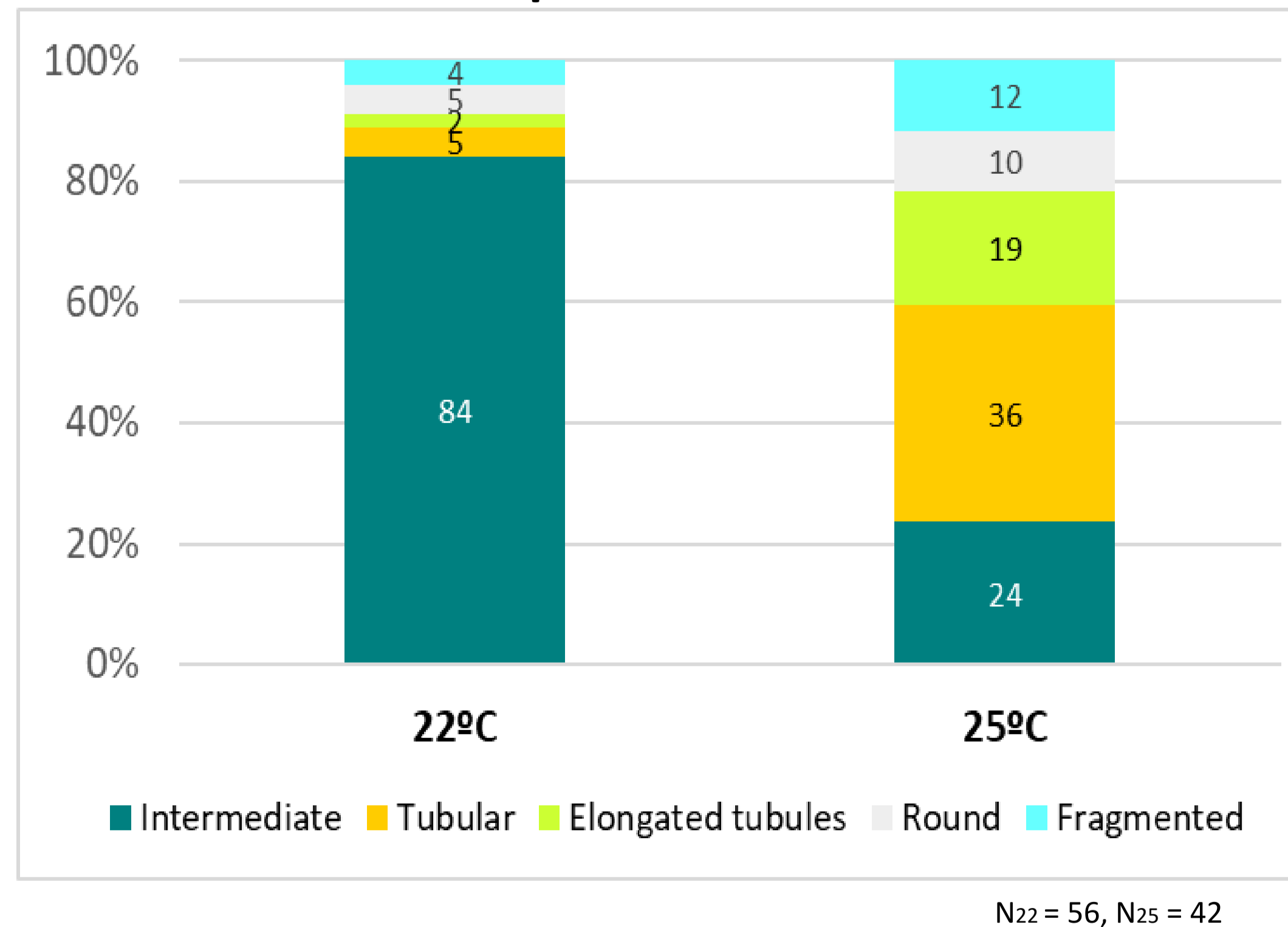


All images were stained with 5 μM Rh123. Color intensity bar for the TPEF signal: dark blue - lowest TPEF signal, dark red - highest TPEF signal. The average laser power in the sample plane - 4-5 mW at 800 nm.

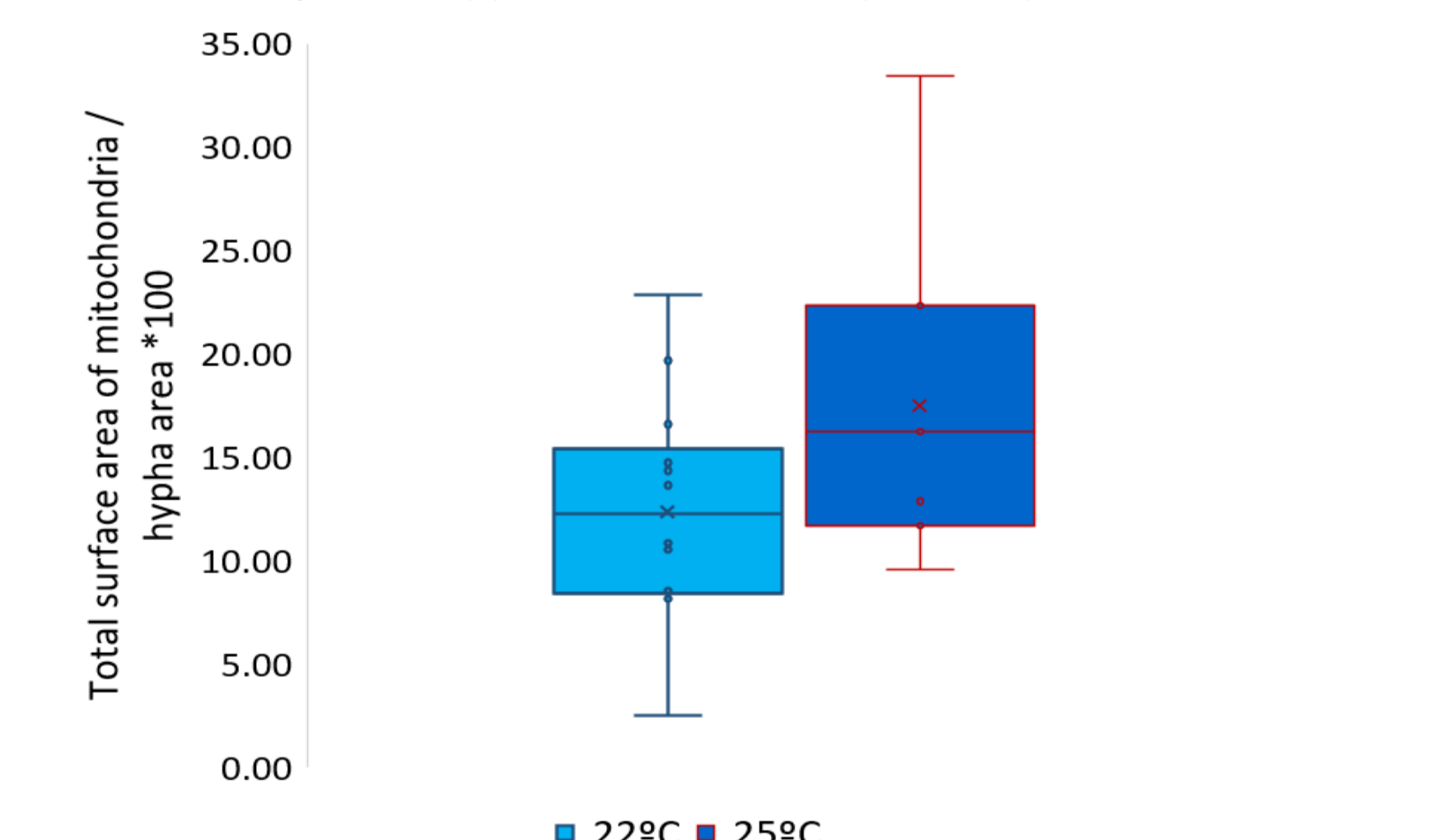
TPEF images analysis for surface area calculation by *Particle Size Analysis* method



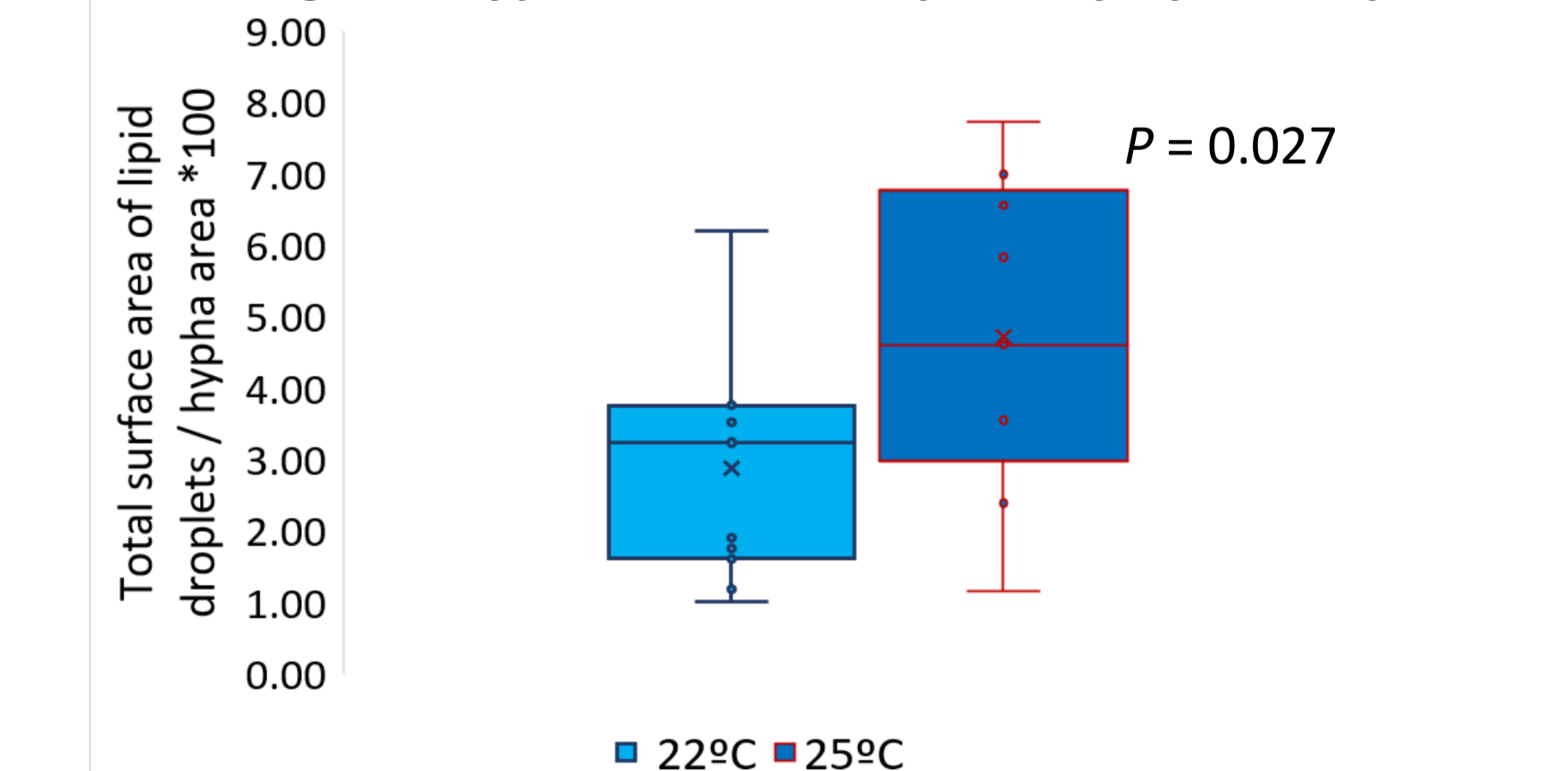
The effect of increased temperature on the abundance of mitochondrial morphology types defined per individual cell



Percentage of hyphal area occupied by mitochondria



Percentage of hyphal area occupied by lipid droplets



Conclusions

Changes in mitochondrial morphology were induced by a small temperature change.

- An increase of 3°C had a dramatic effect on mitochondrial morphology, inducing the appearance of a predominantly tubular morphology.
- The total area percentage of mitochondria showed an increasing trend when grown at 25°C.
- Increasing the ambient temperature to 25°C induced a statistically significant increase in the percentage of hyphal area occupied by LDs from 2.9 ± 1.6 to 4.7 ± 2.2 .

The observed response to the small temperature increase points to the physiological adaptation of hyphal metabolism.

REFERENCES

- [1] K. Ma et al., Front. Cell Dev. Biol. 8, 467 (2020).
- [2] M. Long, T.G. McWilliams, Autophagy 19, 724 (2023).

Acknowledgment

This work was supported by the Ministry of Science, Technological Development and Innovations, Republic of Serbia [contract numbers: 451-03-47/2023-01/200178 and 451-03-47/2023-01/200007]; The Project Advanced Biophysical Methods for Soil Targeted Fungi-Based Biocontrol Agents - BioPhysFUN [Grant number 4545] from Program DEVELOPMENT - Green program of cooperation between science and industry, Science Fund of the Republic of Serbia; the Project HEMMAGINERO [Grant number 6066079] from Program PROMIS, Science Fund of the Republic of Serbia; and the Institute of Physics Belgrade, through the grant by the Ministry of Science, Technological Development and Innovations of the Republic of Serbia.